Synthesis of Adenosine 5'- $[\gamma(R)$ -17O, 18O-thio]triphosphate

Richard C. Bethell and Gordon Lowe*

The Dyson Perrins Laboratory, Oxford University, South Parks Road, Oxford OX1 3QY, U.K.

An efficient synthesis of adenosine $5'-[\gamma(R)^{-17}O,^{18}O-\text{thio}]$ triphosphate has been developed using a combined chemical and enzymic strategy.

Adenosine 5'-triphosphate (ATP) is the universal currency of free energy in living systems and as such is involved in a great many enzyme-catalysed reactions. Stereochemical investigations of many of these reactions have provided important evidence concerning their mechanism of action. Several of the enzymes, however, lead either directly or indirectly to the release of inorganic phosphate and with a view to studying the stereochemical course of such reactions a new method for the configurational analysis of chiral inorganic [^{16}O , ^{17}O , ^{18}O]thiophosphate has recently been developed. We now report an efficient synthesis of adenosine 5'-[$^{7}(R)$ - ^{17}O , ^{18}O -thio]triphosphate which is required as a substrate for these stereochemical investigations.

Several routes to S-nitrobenzyl [¹¹O₃]phosphorothioate (1) were investigated but the method of choice, in terms of overall yield and efficient use of isotopically enriched water, is outlined in Scheme 1. Phosphorus trichloride was hydrolysed quantitatively with a slight excess (1.5 equiv.) of [¹¹O₃]water to [¹¹O₃]phosphorous acid which was converted into its tris-(trimethylsilyl) ester with chlorotrimethylsilane. Bis(S-p-nitrobenzyl)disulphide, prepared by the method of Gladysz et al.,³ on reaction with the PIII triester gave directly S-p-nitrobenzyl [¹¹O₃]phosphorothioate (1). This conversion was essentially quantitative (as determined by ³¹P n.m.r. spectroscopy).

Although adenosine can be phosphorylated selectively at the 5'-hydroxy group by phosphoryl chloride in various solvents,⁴ we and others⁵ have found that 2'- and 3'-substituted adenosine monophosphates (AMP) are also formed (together with small amounts of adenosine bisphosphates) which are difficult to remove. In contrast adenosine is thiophosphorylated exclusively at the 5'-hydroxy group leading after hydrolysis to adenosine 5'-phosphorothioate. By hydrolysing the intermediate with [18O]water, adenosine

PCl₃
$$\xrightarrow{i}$$
 H₃P Φ_3 \xrightarrow{ii} (Me₃Si Φ)₃P
$$\Phi = {}^{17}O$$

$$\rho - O_2N \cdot C_6H_4 \cdot CH_2 \cdot SP\Phi_3$$
(1)

Scheme 1. Reagents: i, [^{17}O]water; ii, Me $_3$ SiCl, Et $_3$ N; iii, (a) (p-O $_2$ N·C $_6$ H $_4$ ·CH $_2$ S $^-$) $_2$, (b) H $_2$ O, NaOH.

Ado
$$\xrightarrow{i}$$
 5'- \bigcirc_2 SP - OAdo \xrightarrow{ii} 5'- \bigcirc_3 P - OAdo \bigcirc = 18 O (2)

Ado = adenosine

Scheme 2. Reagents: i, (a) PSCl₃, (EtO)₃PO; (b) [¹⁸O]water, ii, Br₂, [¹⁸O]water.

5'-[¹⁸O₂]phosphorothioate is formed,⁶ which may be converted into adenosine 5'-[¹⁸O₃]phosphate (2) by treatment with bromine in [¹⁸O]water,⁷ (Scheme 2). This proved to be the most efficient route of several investigated.

Adenosine 5'-[$\gamma(R)$ -1'O,1⁸O-thio]triphosphate was now synthesized from *S-p*-nitrobenzyl [1'O₃]phosphorothioate (1) and adenosine 5'-[1⁸O₃]phosphate (2) as outlined in Scheme 3. Activation of *S-p*-nitrobenzyl [1'O₃]phosphorothioate (1) with diphenylphosphoryl chloride, followed by coupling with adenosine 5'-[1⁸O₃]phosphate using a modification of the Michelson procedure, gave after reductive cleavage of the *S*-nitrobenzyl group with sodium in liquid ammonia, adenosine 5'-[α -1⁸O₂, α β -1⁸O, β -1⁷O₂-thio]diphosphate (3). A preliminary study of the coupling of *S*-benzyl [1⁸O₃]phosphorothioate with adenosine 5'-phosphate had established that the bridge oxygen is derived exclusively from AMP.

Pyruvate kinase phosphorylates adenosine 5'-β-thiodiphosphate (ADPβS) with phosphoenolpyruvate to give predominantly adenosine 5'-[(S)-β-thio]triphosphate [(S_p)-ATPβS] (ca. 80%) and some (R_p)-ATPβS (ca. 20%). The (R_p)-ATPβS, however, is the diastereoisomer overwhelmingly preferred by yeast hexokinase and so by incubating the diastereoisomers with hexokinase and D-glucose, (S_p)-ATPβS and ADPβS are obtained which are readily separated. Incubation of adenosine 5'-[α -18O₂, α β-18O,β-17O₂-thio]diphosphate (3) with pyruvate kinase and phosphoenolpyruvate gave a mixture of (S_p)-[17O₂,18O₃]ATPβS and (R_p)-[17O₂,18O₃]ATPβS which after incubation with hexokinase

Scheme 3. Reagents: i, (a) (PhO)₂POCl, (b) [¹⁸O₃]AMP; ii, Na, liquid NH₃; iii, (a) pyruvate kinase, phosphoenolpyruvate, (b) hexokinase, p-glucose; iv, methionyl-tRNA synthetase, methionine.

and D-glucose gave (S_p) -[$^{17}O_2$, $^{18}O_3$]ATP β S (4). The recovered [$^{17}O_2$, $^{18}O_3$]ADP β S was recycled using the combined action of pyruvate kinase (with phosphoenolpyruvate) and hexokinase (with D-glucose). By this means adenosine 5'-[α - $^{18}O_2$, $\alpha\beta$ - $^{18}O_3$ - $^$

Most aminoacyl-tRNA synthetases will catalyse the activation of their specific amino acid in the absence of the cognate tRNA. The activating agent is MgATP and the product is the aminoacyl-adenylate and Mg inorganic pyrophosphate: the process is reversible.11 Methionyl-tRNA synthetase accepts (S_n) -ATP β S as a substrate and in the presence of methionine converts it into adenosine 5'-(y-thio)triphosphate (ATPyS), via the methionyl adenylate, this being the thermodynamically more stable nucleotide. 12 Incubation of adenosine 5'- $[\alpha$ - $^{18}O_2$, $\alpha\beta^{-18}O$, $\beta(S)^{-17}O_2$ -thio] triphosphate (4) with methionine and methionyl-tRNA synthetase from Bacillus stearothermophilus gave adenosine 5'- $[\alpha^{-18}O_2,\beta\gamma^{-17}O,\gamma(R)$ -¹⁷O, ¹⁸O-thioltriphosphate (5), referred to simply as adenosine 5'- $[\gamma(R)^{-17}O, ^{18}O$ -thio]triphosphate in the title. The isolated yield on this enzyme-catalysed rearrangement was 76%. Adenosine 5'- $[\gamma(S)^{-17}O, ^{18}O$ -thio]triphosphate has been synthesized previously, 13 but the incorporation of [17O] water was much less efficient making the synthesis expensive if carried out on a substantial scale; moreover, difficulties in repeating the synthesis have been encountered. 14 The synthesis reported here started with 2 mmol of phosphorus trichloride which was hydrolysed with 9 mmol of [17O] water (171 mg) to give 0.32 mmol of adenosine 5'- $[\gamma(R)$ -17O, 18O-thio]triphosphate (5).

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References

- 1 G. Lowe, Acc. Chem. Res., 1983, 16, 244; F. Eckstein, Annu. Rev. Biochem., 1985, 54, 367.
- 2 J. R. P. Arnold and G. Lowe, J. Chem. Soc., Chem. Commun., 1986, 865.
- 3 J. A. Gladysz, V. K. Wong, and B. S. Jick, *Tetrahedron*, 1979, 35, 2329.
- 4 M. Yoshikawa, T. Kato, and T. Takenishi, Bull. Chem. Soc. Jpn., 1969, 42, 3505; T. Sowa and S. Ouchi, ibid., 1975, 48, 2084.
- 5 W. H. Dawson, R. L. Cargill, and R. B. Dunlap, J. Carbohydr. Nucleosides Nucleotides, 1977, 4, 363.
- 6 J. P. Richard and P. A. Frey, J. Am. Chem. Soc., 1982, 104, 3476.
- 7 G. Lowe, G. Tansley, and P. M. Cullis, J. Chem. Soc., Chem. Commun., 1982, 595; G Lowe, B. S. Sproat, G. Tansley, and P. M. Cullis, Biochemistry, 1983, 22, 1229.
- 8 A. M. Michelson, Biochim. Biophys. Acta, 1964, 91, 1.
- 9 E. K. Jaffe and M. Cohn, J. Biol. Chem., 1978, 253, 4823.
- 10 D. Yee, V. W. Armstrong, and F. Eckstein, Biochemistry, 1979, 18, 4116
- 11 P. R. Schimmel and D. Söll, Annu. Rev. Biochem., 1979, 48, 601.
- 12 E. F. Rossomando, L. T. Smith, and M. Cohn, *Biochemistry*, 1979, 18, 5670.
- M. R. Webb and D. R. Trentham, J. Biol. Chem., 1981, 256, 4884;
 M. R. Webb, Methods Enzymol., 1982, 87, 301.
- 14 P. A. Frey, personal communication.